

Porphyromonas gingivalis and Alzheimer's disease: A narrative review

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Abstract

Objective: The purpose of this review is to address the purported association of periodontitis and the microbial pathogen, *Porphyromonas gingivalis* (*P. gingivalis*), with Alzheimer's disease (AD).

Methods: A scoping search was conducted using terms related to biological plausibility, comorbid associations of periodontitis, *P. gingivalis* and AD. Ovid Medline and PubMed were used to obtain abstracts which, in turn, were used to select papers for further evaluation. Special attention was given to articles addressing confounders, risk factors, and causality. A total of 113 papers from the basic science and medical literature and 48 papers from the dental literature, covering the years 2010 through May 2025 were reviewed.

Results: Evidence exists showing that inflammation associated with periodontitis and *P. gingivalis* infection has implications for brain health. There is evidence to establish biologic plausibility for a microbial role in promoting neuroinflammation and neurodegeneration as seen in AD.

Conclusion: There is evidence that both *P. gingivalis* and periodontitis satisfy the requirements of biological plausibility, have comorbid relationships, and may be risk factors for AD. However, there is insufficient evidence to establish *P. gingivalis* or periodontitis as independent causes of AD.

Keywords: *Porphyromonas gingivalis*. Alzheimer's Disease. Chronic periodontitis. Risk factors. Virulence factors.

Introduction

Severe periodontitis has a high prevalence, affecting 11.2% of the world's population making it the sixth most common human disease¹. Even in developed countries, such as the United States (U.S.), the prevalence of severe periodontitis can be significant, i.e., 7.8% for adults, age 30 to 79 years². Based on the latest U.S. census data, this prevalence of disease extrapolates to roughly 6.5 million people³.

A significant body of literature is supportive of independent associations between severe periodontitis and a variety of systemic diseases, such as atherosclerosis^{4,5}, cardiovascular disease^{1,6}, stroke⁷⁻¹⁰, and Alzheimer's disease¹¹⁻¹⁴. Thus, the purpose of the present narrative review is to address the purported association of periodontitis and, more specifically, of *Porphyromonas gingivalis* (*P. gingivalis*), with Alzheimer's disease (AD).

Methods

A scoping search was conducted using terms related to biological plausibility, comorbid associations of both periodontitis and AD, and virulence factors associated with *P. gingivalis*. Ovid Medline, PubMed and the reference sections of published papers were used to obtain abstracts which, in turn, were used to select papers for further evaluation. Special attention was given to articles addressing confounders, risk factors, and causality. A total of 113 papers from the basic science and medical literature, and 48 papers from the dental literature, covering the years 2010 through May 2025, were reviewed.

Relevant terminology

Assessment of the literature pertaining to periodontitis and its association with various systemic diseases requires the reader to differentiate between biologic plausibility, causality, conflation, risk factors, comorbidity, and confounders.

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Biological plausibility involves rational thought in the evaluation of a possible cause-and-effect relationship between a biological factor and a particular disease. Evidence-based criteria for treatment of a disease are ultimately based on biological plausibility, as the concept helps in assessing the strength of evidence and associations observed in epidemiological studies^{15,16}.

Causality, in simple terms, is the relationship between cause and effect. A more sophisticated definition is that causation refers to a process in which an initial event induces the occurrence of a subsequent event¹⁷. In the pathobiology realm, causality is difficult to prove¹⁸. Studies proving causality require significant assumptions, in-depth subject-matter knowledge, precise statistical analysis, and consideration of alternative explanations¹⁹. Interestingly, in 2017, the World Health Organization proclaimed the term causal association “*is only used if there is no other factor intervening in the processes*”²⁰. Any discussion of causality is incomplete without noting the contributions of the Bradford-Hill criteria²¹. The Bradford Hill criteria provide a framework for moving from an observed association to a conclusion of causation. The nine criteria are: strength of association between a risk factor and outcome; consistency of findings; specificity of association, i.e., a one-to-one relationship between risk factor and outcome; temporality sequence or exposure must precede outcome; a dose-response gradient; biologic plausibility or a feasible biologic mechanism; coherence between epidemiological and laboratory findings; experimental evidence; and similarities between the observed association and any other associations or an analogous event²¹.

Conflation can be defined as the erroneous fusion or blending of two similar but disparate concepts into one, which in medical research is often referred to as “being equal to”²². Conflation is more commonly noted in observational studies, and generally results in biased estimates or conclusions²².

With respect to disease, risk factors are defined as a characteristics that are associated with an increased prevalence of a disease²³. In the case of periodontal disease, most risk factors are modifiable and include poor oral hygiene, tobacco use, alcohol consumption, psychological stress, depression, osteoporosis, and metabolic syndrome^{24,25}. Examples of non-modifiable risk factors are genetic susceptibility, and age²³⁻²⁵.

Comorbidity simply refers to the simultaneous presence of two or more medical conditions^{26,27}. As previously noted, there are associations between periodontitis and a variety of comorbid systemic diseases. For many of these associations, shared inflammatory pathways appear to be the common contributing factor²⁷.

In research designed to determine a potential cause-and-effect relationship, a confounding variable is an unmeasured third factor that may influence either the cause

or the effect, or both. It's important to consider potential confounding variables and account for them in clinical trials, to ensure that treatment outcomes are valid. Unidentified or unaccounted for confounders can introduce bias and/or cause an invalid interpretation of results. In epidemiological studies of periodontal disease, it appears that age, gender and race are the most frequently included confounders^{28,29}.

Epidemiology of Alzheimer's disease

AD is the most common form of dementia and one of the leading sources of morbidity and mortality in the elderly. The incidence and prevalence of AD is reportedly increasing but likely underestimated, due to underdiagnosis and misdiagnosis^{30,31}. The global prevalence of dementia was estimated to be 57.4 (95% uncertainty interval 50.4 – 65.1) million in 2019, and was predicted to be more than doubled over the next 30 years to 2050³¹. Li et al.³² have noted that between the years 1990 and 2019, the incidence and prevalence of AD and other dementias increased by 147.95% and 160.84%, respectively. The authors also reported that smoking was a major risk factor in men, while obesity was the major risk factor for women³².

Etiology of Alzheimer's disease

Despite improved understanding of the pathogenesis of AD, the etiology remains unclear. Current treatment protocols do not prevent the disease and only marginally impact progression³³⁻³⁵. AD may be considered a multifactorial disease with multiple risk factors, such as increasing age, genetics, vascular disease, head trauma, environmental factors, and infection^{33,35,36}.

There is ample evidence that pathogens within the oral microbiome can be a factor in cerebrovascular and neurodegenerative pathology³⁷⁻⁴⁰. Invasive pathogenic bacteria can exhibit a variety of immunomodulatory functions, including the activation of systemic pro-inflammatory cytokines⁴¹. Interestingly, amyloid- β has been shown to express antiviral and antimicrobial activity, a characteristic that may help explain its accumulation in brain parenchymal tissues that have experienced bacterial infection^{36,42-44}.

The neuropathologic changes in AD include disseminated neuritic plaques with discernible extracellular beta-amyloid deposition, dystrophic neurites, and neurofibrillary tangles that include intracellular accumulation of tau protein^{45,46}. Additionally, these features are associated with activated astrocytes and the proliferation and activation of microglial cells. Microglia cells are the resident macrophages of the central nervous system and function in much the same manner as their more peripheral counterparts⁴⁷. Both astrocytes and activated microglia cells contribute to neuroinflammation

and neurodegeneration phenomena⁴⁸⁻⁵². Indeed, a recent study has reported that microglia cells in the brains of people with Alzheimer's disease were more likely to be in a pre-inflammatory state and more likely to produce inflammatory mediators that damage brain cells⁵³.

Periodontitis and Alzheimer's disease

Stage III and IV periodontitis⁵⁴ and AD are relatively common comorbid conditions in the elderly^{2,14,26,55-58}. Several papers have noted that periodontitis is associated with increased severity of dementia and a more rapid cognitive decline in AD⁵⁵⁻⁶⁰. Interestingly, as noted previously, the number of U.S. adults between the ages of 30 and 75 years afflicted with severe periodontitis is reported at 7.8%, or approximately 6.5 million adults². The Alzheimer's Association reports that 6.9 million U.S. adults, age ≥ 65 years, have AD, which extrapolates to roughly 7% of the population⁶¹.

Porphyromonas gingivalis

Contemporary opinion holds that periodontitis is an inflammatory disease involving a community of bacteria in which various communal species are in a dysbiotic state⁶²⁻⁶⁵. The dysbiosis, when combined with the host immune response, favors establishment of chronic inflammation in the periodontal tissues^{65,66}. If not treated, or if inadequately treated, the host inflammatory response persists and initiates a degradation of the tooth-supporting tissues and, in advanced stages, may lead to loss of teeth⁶⁷. The biofilm associated with periodontitis exhibits a wonderfully complex microbial community (Fig. 1), which is predominately anaerobic, Gram-negative, and proteolytic. An often cited offender and initiator of dysbiosis in the biofilm associated with periodontitis is *P. gingivalis*^{62,66,67}.

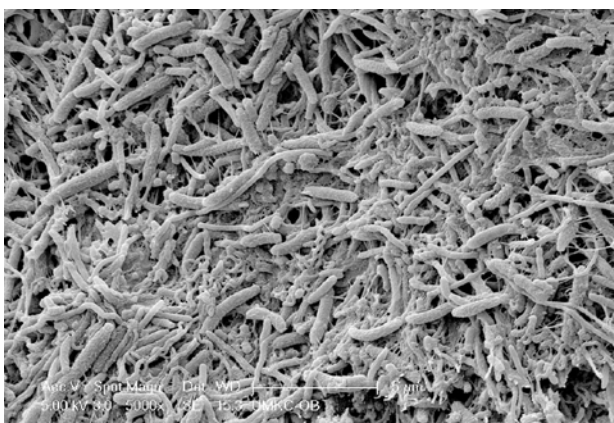


Figure 1. Scanning electron microscopy view of a complex, mature biofilm on the root of a tooth extracted due to advanced periodontitis (stage IV). Identifiable morphotypes of bacteria include long, short and curved rods of various diameters, fusiforms, spirochetes, and cocci. Original magnification of 5,000x. Bar = 5 μ m.

P. gingivalis is a short rod-shaped, Gram negative, anaerobic, asaccharolytic bacterium that forms black-brown colonies on enriched blood agar. *P. gingivalis* has been detected in $\approx 86\%$ of subgingival plaque samples from periodontitis patients⁶⁸. The microbe is a critical component in the pathogenesis and progression of periodontitis. Indeed, it has been characterized as a 'keystone' pathogen whose association with other bacteria can dictate the conversion of a biofilm composed of commensal microorganisms into a complex dysbiotic biofilm that interacts in an adversarial manner with the host immune system, which, in turn, results in a tissue destructive inflammatory response, i.e., periodontitis⁶⁸⁻⁷¹.

P. gingivalis possesses a number of virulence factors that help define its' pathogeny^{68,72,73}. For example, the virulence factors of *P. gingivalis* play important roles in microbial coaggregation, biofilm formation, oral biofilm dysbiosis, and the pathogenesis of periodontitis. The primary virulence factors of *P. gingivalis* include:

- 1) Lipopolysaccharide (LPS), a major component of the outer cell membrane, with a well-established ability to cause inflammation^{74,75}.
- 2) Fimbriae, which are filamentous surface appendages, extending from the outer cell membrane (Fig. 2), that function in bacterial motility, adhesion to host cells, and facilitate invasion into host cells and tissues^{76,77}.
- 3) The capsule, which, in the case of *P. gingivalis*, serves multiple functions. For example, the capsule protects against phagocytosis by host leukocytes, facilitates co-aggregation with other bacteria in the assembly of a biofilm mass, and can induce host immune responses, such as cytokine expression by dendritic cells and chemokine expression by macrophages^{67,73,78,79}.
- 4) Outer membrane vesicles (OMVs), produced via blebbing of prokaryotic (bacteria) membranes (Fig. 2), have frequently been identified as cell debris or artifacts induced

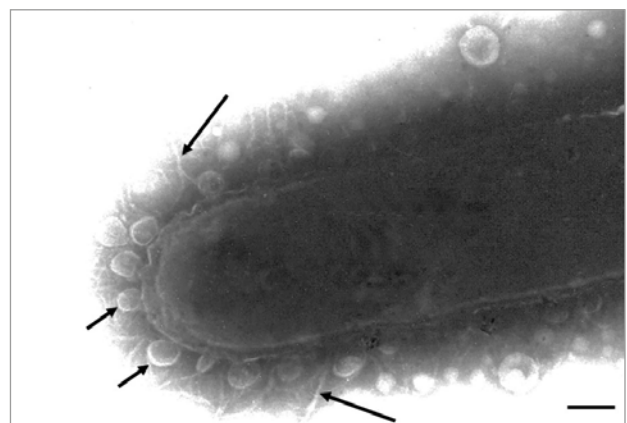


Figure 2. Transmission electron microscopy view of unsectioned, negative-stained *P. gingivalis* (ATCC 33277), demonstrating outer membrane vesicles (short arrow) and fimbria (long arrow). Stained with 2% potassium phosphotungstate. Original magnification of 5,000x. Bar = 0.05 μ m.

by processing for electron microscopy. However, research on OMVs has shown them to be involved in the transport of molecular effectors, cell-cell interactions, nutrients, host cell immune modulation and dysregulation, bacterial aggregation, and biofilm formation⁸⁰. OMVs of *P. gingivalis* are reported to contain a toxic complex of LPS, proteolytic enzymes, gingipain, and pathogen-derived DNA and RNA⁸¹⁻⁸³.

5) Gingipain is a family of cysteine proteinases that, depending on the strain of *P. gingivalis*, can either be anchored on the bacterial membrane or released via OMVs into the extracellular environment⁸⁴. Gingipains are potent virulence factors and account for 85% of extracellular proteolytic activity of *P. gingivalis*. Gingipains dysregulate the host immune response and promote inflammation, inactivation of immune inhibitors, and cleavage of immune cell receptors⁸⁵. In addition, gingipains can degrade macrophage CD14 and thereby avoid phagocytosis by host leukocytes and induce loss of viability in host fibroblasts and endothelial cells^{86,87}. Gingipains have also been implicated in the activation of *P. gingivalis* collagenase and degradation of type I collagen⁸⁸. *In vitro* studies have shown *P. gingivalis* capable of increasing collagen degradation through a combination of increased activation of human gingival fibroblast matrix metalloproteinase (MMP) and decreasing levels of tissue inhibitor of metalloproteinase-1 (TIMP-1)^{89,90}. *P. gingivalis* also up-regulates mRNA expression of multiple MMPs and TIMPs⁸⁹.

6) *Porphyromonas gingivalis* can invade epithelial cells and other periodontal tissue cells, and thus evade host antimicrobial defenses. In doing so, the microbe utilizes a variety of virulence factors that cause deregulation of the host innate immune and inflammatory responses. For example, *P. gingivalis* has been shown to subvert the response of neutrophils, the most efficient phagocyte in the inflammatory process, by utilizing gingipains, serine proteases, lipid phosphatases, lipopolysaccharide, and fimbriae^{85,91}. Disruption of the host immune response allows for opportunistic tissue invasion and promotes inflammation and tissue damage⁷³.

***Porphyromonas gingivalis* and Alzheimer's disease**

Even a cursory review of current literature reveals that inflammation is a major component in the pathogenesis of neurodegeneration seen in AD and possibly other forms of dementia. The role of neuroinflammation in AD raises the question of whether chronic inflammatory conditions or inflammation promoting factors peripheral to the central nervous system (CNS) contribute to the CNS inflammatory burden.

The role attributed to *P. gingivalis* in periodontitis almost makes it a surrogate microbe for the disease. Clearly *P. gingivalis* is not a singular microbial

pathogen in the cause and progression of periodontitis. Certainly, other pathogens and their panel of virulence factors play critical roles in periodontitis and, indirectly, in the promotion of systemic inflammation, and potentially in AD⁹²⁻⁹⁵. Multiple studies have established that elevated antibodies to periodontal bacteria are associated with an increased systemic pro-inflammatory state; and that increased levels of pro-inflammatory cytokines have been associated with an increased rate of cognitive decline in Alzheimer's disease^{55,57,96-98}.

As previously noted, *P. gingivalis* is just one of several bacterial pathogens that have been associated with AD in the research literature. However, in terms of pathogenicity, prominent role in severe periodontitis, generation of peripheral inflammation, and ability to invade the CNS, *P. gingivalis* has become the target microbe for studies designed to determine the biological plausibility and potential role as risk factors of microbes in AD. Consider, for example, depending on serotype, that *P. gingivalis* can be capsulated or non-capsulated, and both are highly pathogenic. The non-capsulated *P. gingivalis* can generate neurodegenerative AD-like changes *in vitro*; and a capsulated *P. gingivalis* has been shown to produce the cardinal hallmark lesions of AD in an animal model and *in vitro*^{99,100}. All *P. gingivalis* serotypes express proteases that enable cleavage of the precursor of amyloid- and tau protein, resulting in the formation of amyloid- and neurofibrillary tangles. Tau is a substrate for gingipains that cleave tau into various peptides. *P. gingivalis* related inflammation has metabolic implications for brain health, as the microbe has been isolated from both atherosclerotic plaque¹⁰¹ and the brains of AD patients¹⁰²⁻¹⁰⁶. The finding of microbes in brain tissue verifies the invasiveness of the organisms and supports the hypothesis for a microbial role in promoting neuroinflammation and neurodegeneration as seen in AD¹⁰⁷⁻¹¹¹.

The pathways for microbial access to the brain include, among several, a damaged blood-brain barrier or through a nerve trunk^{19,107}. *P. gingivalis* can trigger both peripheral and cerebral immune responses¹⁰⁸ and, when combined with defective susceptibility genes (such as apolipoprotein E4 and complement receptor 1), can prime microglial cells into the pro-inflammatory phenotype¹¹² often associated with AD.

When the aggregate of published evidence is evaluated, there is a compelling argument supporting the biological plausibility for *P. gingivalis* as a comorbidity or conceivably a risk factor in AD pathophysiology (Fig. 3). However, there is insufficient evidence indicating a causal role for severe periodontitis or *P. gingivalis* infection. Clearly, there is no universally accepted answer to the question: Does improvement in periodontal health lower the risk of developing Alzheimer's disease?

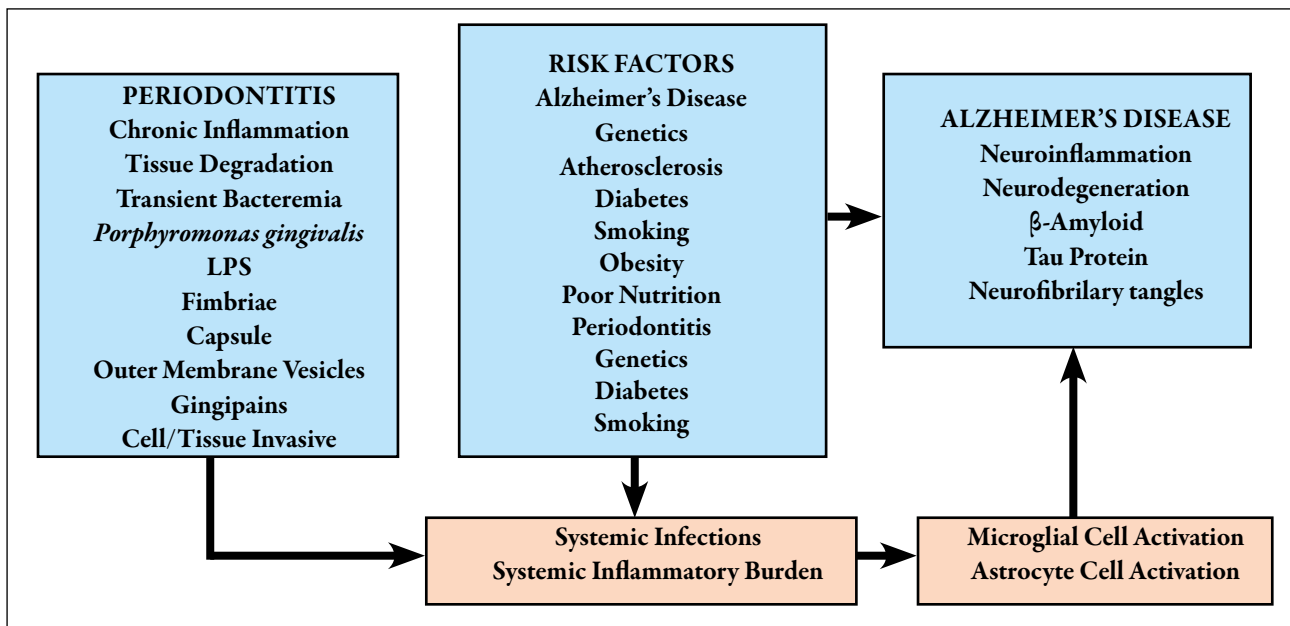


Figure 3. Biological plausibility of the relationship between periodontitis and Alzheimer's disease.

Caveat

To paraphrase Abt et al.¹¹³, when confronted with numerous clinical studies, all reaching the conclusion that periodontitis is 'associated' with AD, how does one judge the importance of such studies? The studies generally use similar designs, share similar limitations and reach similar conclusions. The clinician must be aware of comorbid conditions, confounding factors, and conflation in such studies. And most importantly, the concept that association with a specific disease or a correlational finding does not mean causality¹¹⁴.

Conclusions

Although there is no evidence establishing periodontitis or *P. gingivalis* as causative of AD, inflammation is an important host response in the comorbid conditions of periodontitis and Alzheimer's disease (AD). Further, there is evidence that both *P. gingivalis* and periodontitis may be risk factors for AD. Given the role of *P. gingivalis* in stages III and IV periodontitis and the role of periodontitis as a comorbid condition with AD, it seems reasonable that physicians should include a visual examination of the teeth and gingivae as part of every patient's physical assessment, and that dentists and dental hygienists should conduct a complete periodontal examination when providing routine patient care^{115,116}. To do less is a disservice to the patient.

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Writing the article: CMC, DES.

Critical revision of the article: CMC, DES, P-CL, KMS, SRM.

Final approval of the article: CMC, DES, P-CL, KMS, SRM.

Overall responsibility: CMC, DES.

References

1. Sanz M, Del Castillo AM, Jepsen S, Gonzalez-Juanatey J, D'Aiuto F, Bouchard P, et al., Periodontitis and cardiovascular diseases: consensus report. *J Clin Periodontol*. 2020 March;47(3):268-288. doi: 10.1111/jcpe.13189.
2. Eke PI, Borgnakke WS, Genco RJ. Recent epidemiologic trends in periodontitis in the USA. *Periodontol* 2000. 2020 Feb;82(1):257-267. doi: 10.1111/prd.12323.
3. United States Census Bureau. 2024 Census Survey. Washington (DC): U.S. Census Bureau; 2024 [Access 25 Feb 2025]. Available from: <https://www.census.gov>.
4. Dong W, Gong Y, Yang B, Lie B. Bioinformatics analysis reveal potential crosstalk genetic and immune relationships between atherosclerosis and periodontitis. *Sci Rep*. 2023 June;13(1):10381. doi: 10.1038/s41598-023-37027-x.
5. Zhou LJ, Lin WZ, Meng XQ, Zhu H, Liu T, Du LJ, et al. Periodontitis exacerbates atherosclerosis through *Fusobacterium nucleatum*-promoted hepatic glycolysis and lipogenesis. *Cardiovas Res*. 2023 Jul;119(8):1706-1717. doi: 10.1093/cvr/cvad045.
6. Herrera D, Sanz M, Shapira L, Brotons C, Chapple I, Frese T, et al. Periodontal diseases and cardiovascular diseases, diabetes, and respiratory diseases: summary of the consensus report by the European Federation of Periodontology and WONCA Europe. *Eur J Gen Pract*. 2024 Dec;30(1):2320120. doi: 10.1080/13814788.2024.2320120.
7. Freiherr Von Seckendorff A, Nomenjanahary MS, Labreuche J, Ollivier V, Di Meglio L, Dupont S, et al. Periodontitis in ischemic stroke: impact of *Porphyromonas gingivalis* on thrombus composition and ischemic stroke outcomes. *Res Pract Thromb Haemost*. 2024 Jan;8(1):102313. doi: 10.1016/j.rpth.2023.102313.
8. Zheng X, Li X, Zhen J, Xue D, Hu J, Cao Q, et al. Periodontitis is associated with stroke. *J Transl Med*. 2023 Oct;21(1):697. doi: 10.1186/s12967-023-04545-1.
9. Ma C, Wu M, Gao J, Liu C, Xie Y, Lv Q, et al. Periodontitis and stroke: a Mendelian randomization study. *Brain Behav*. 2023 Feb;13(2):e2888. doi: 10.1002/brb3.2888.
10. Almarhoumi R, Alvarez C, Harris T, Tognoni CM, Paster BJ, Carreras I, et al. Microglial cell response to experimental periodontal disease. *J Neuroinflammation*. 2023 June;20(1):142. doi: 10.1186/s12974-023-02821-x.
11. Lee YT, Tsai CF, Yen YC, Huang LK, Chao SP, Hu LY, et al. Periodontitis is a potential risk factor for transient ischemic attack and minor ischemic stroke in young adults: a nationwide population-based cohort study. *J Periodontol*. 2022 Dec;93(12):1848-1856. doi: 10.1002/JPER.21-0528.
12. Wu H, Qiu W, Zhu X, Li X, Xie Z, Carreras I, et al. The periodontal pathogen *Fusobacterium nucleatum* exacerbates Alzheimer's pathogenesis via specific pathways. *Front Aging Neurosci*. 2022 Jun;14:912709. doi: 10.3389/fnagi.2022.912709.
13. Dibello V, Lozupone M, Manfredini D, Dibello A, Zupo R, Sardone R, et al. Oral frailty and neurodegeneration in Alzheimer's disease. *Neural Regen Res*. 2021 Nov;16(11):2149-2153. doi: 10.4103/1673-5374.310672.
14. Werber T, Bata Z, Vaszine ES, Berente DB, Kamondi A, Horvath AA. The association of periodontitis and Alzheimer's disease: how to hit two birds with one stone. *J Alzheimer's Dis*. 2021;84(1):1-21. doi: 10.3233/JAD-210491.
15. Dailey J, Rosman L, Silbergeld EK. Evaluating biological plausibility in supporting evidence for action through systematic reviews in public health. *Public Health*. 2018 Dec;165:48-57. doi: 10.1016/j.puhe.2018.08.015.

16. Whaley P, Piggott T, Morgan RL, Hoffmann S, Tsaioun K, Schwingshackl L, et al. Biological plausibility in environmental health systematic reviews: a GRADE concept paper. *J Clin Epidemiol*. 2022 Jun;146:32-46. doi: 10.1016/j.jclinepi.2022.02.011.
17. Hernán MA, Robins JM. Estimating causal effects from epidemiological data. *J Epidemiol Community Health*. 2006 Jul;60(7):578-586. doi: 10.1136/jech.2004.029496.
18. Rothman KJ, Greenland S. Causation and causal inference in epidemiology. *Am J Public Health*. 2005 Jul;95 Suppl 1:S144-S150. doi: 10.2105/AJPH.2004.059204.
19. Joffe M, Gambhir M, Chadeau-Hyam M, Vineis P. Causal diagrams in systems epidemiology. *Emerg Themes Epidemiol*. 2012 Mar;9(1):1. doi: 10.1186/1742-7622-9-1.
20. Puliyl J, Naik P. Revised World Health Organization (WHO)'s causality assessment of adverse events following immunization – a critique. *F1000Res*. 2018 Feb;7:243. doi: 10.12688/f1000research.13694.2.
21. Hill AB. The environment and disease: association or causation? *Proc R Soc Med*. 1965 May;58(5):295-300. doi: 10.1177/003591576505800503.
22. Ramspek CL, Steyerberg EW, Riley RD, Rosendaal FR, Dekkers OM, Dekker FW, et al. Prediction or causality? A scoping review of their conflation within current observational research. *Eur J Epidemiol*. 2021 Sept;36(9):889-898. doi: 10.1007/s10654-021-00794-w.
23. Van Dyke TE, Sheilesh D. Risk factors for periodontitis. *J Inter Acad Periodontol*. 2005 Jan;7(1):3-7.
24. Reynolds MA. Modifiable risk factors in periodontitis: at the intersection of aging and disease. *Periodontol* 2000. 2014 Feb;64(1):7-19. doi: 10.1111/prd.12047.
25. Genco RJ, Borgnakke WS. Risk factors for periodontal disease. *Periodontol* 2000. 2013 June;62(1):59-94. doi: 10.1111/j.1600-0757.2012.00457.x.
26. Hajishengallis G. Interconnection of periodontal disease and comorbidities: evidence, mechanisms, and implications. *Periodontol* 2000. 2022 June;89(1):9-18. doi: 10.1111/prd.12430.
27. Holmstrup P, Damgaard C, Olsen I, Klinge B, Flyvbjerg A, Nielsen CH, et al., Comorbidity of periodontal disease: two sides of the same coin? An introduction for the clinician. *J Oral Microbiol*. 2017 June;9(1):1332710. doi: 10.1080/20002297.2017.1332710.
28. Natto ZS, Ahmad RHA, Alsharif LT, Alrowithi HF, Alsini DA, Salih HA, et al., Chronic periodontitis case definitions and confounders in periodontal research: a systematic assessment. *BioMed Res Int*. 2018 Nov;2018:4578782. doi: 10.1155/2018/4578782.
29. Safiri S, Jolfayi AG, Fazlollahi A, Morsali S, Sarkesh A, Sorkhabi AD, et al. Alzheimer's disease: a comprehensive review of epidemiology, risk factors, symptoms diagnosis, management, caregiving, advanced treatments and associated challenges. *Front Med*. (Lausanne) 2024 Dec;11:1474043. doi: 10.3389/fmed.2024.1474043.
30. Monfared AAT, Byrnes MJ, White LA, Zhang Q. Alzheimer's disease: epidemiology and progression. *Neurol Ther*. 2022 June;11(2):553-569. doi: 10.1007/s40120-022-00338-8.
31. GBD 2019 Dementia Forecasting Collaborators. Estimation of the global prevalence of dementia in 2019 and forecasted prevalence in 2050: an analysis for the global burden of disease study 2019. *Lancet Public Health*. 2022 Feb;7(2):e105-e125. doi: 10.1016/S2468-2667(21)00249-8.
32. Li X, Feng X, Sun X, Hou N, Han F, Liu Y. Global, regional, and national burden of Alzheimer's disease and other dementias, 1990-2019. *Front Aging Neurosci*. 2022 Oct;14:937486. doi: 10.3389/fnagi.2022.937486.
33. Breijyeh Z, Karaman R. Comprehensive review on Alzheimer's disease: causes and treatment. *Molecules*. 2020 Dec;25(24):5789. doi: 10.3390/molecules25245789.
34. Passeri E, Elkhoury K, Morsink M, Broersen K, Linder M, Tamayol A, et al. Alzheimer's disease: treatment strategies and their limitations. *Inter J Mol Sci*. 2022 Nov;23(22):13954. doi: 10.3390/ijms232213954.
35. Uleman JF, Melis RJE, Hoekstra AG, Olde Rikkert MGM, Quax R, Australian Imaging, Biomarker and Lifestyle Study of Aging and Alzheimer's Disease Neuroimaging Initiative Studies. Exploring the potential impact of multi-factor precision interventions in Alzheimer's disease with system dynamics. *J Biomed Inform*. 2023 Sept;145:104462. doi: 10.1016/j.jbi.2023.104462.
36. Loughman A, Adler CJ, Macpherson H. Unlocking modifiable risk factors for Alzheimer's disease: does the oral microbiome hold some of the keys? *J Alzheimer's Dis*. 2023 Apr;92(4):1111-1129. doi: 10.3233/JAD-220760.
37. Aoki S, Nishi H, Shiga Y, Nezu T, Eto F, Imamura E, et al. *Fusobacterium nucleatum* in the oral cavity is associated with cerebral small vessel disease in patients with ischemic stroke. *J Stroke Cerebrovasc Dis*. 2025 Jan;34(1):108183. doi: 10.1016/j.jstrokecerebrovasdis.2024.108183.
38. Freiherr Von Seckendorff A, Nomenjanahary MS, Labreuche J, Ollivier V, Di Meglio L, Dupont S, et al. Periodontitis in ischemic stroke: impact of *Porphyromonas gingivalis* on thrombus composition and ischemic stroke outcomes. *Res Pract Thromb Haemost*. 2024 Jan;8(1):102313. doi: 10.1016/j.rpth.2023.102313.
39. Jang KA, Kim YR, Joo K, Son M. Chronic periodontitis and risk of cerebro-cardiovascular diseases among older Koreans. *Gerodontology*. 2024 Sept;41(3):400-408. doi: 10.1111/ger.12722.
40. Leonov G, Salikhova D, Starodubova A. Oral microbiome dysbiosis as a risk factor for stroke: A comprehensive review. *Microorganisms*. 2024 Aug;12(8):1732. doi: 10.3390/microorganisms12081732.
41. Baker JL, Mark Welch JL, Kauffman KM, McLean JS, He X. The oral microbiome: diversity, biogeography and human health. *Nat Rev Microbiol*. 2024 Feb;22(2):89-104. doi: 10.1038/s41579-023-00963-6.
42. Vidasova D, Nemergut M, Liskova B, Damborsky J. Multi-pathogen infections and Alzheimer's disease. *Microb Cell Fact*. 2021 Jan;20(1):25. doi: 10.1186/s12934-021-01520-7.

43. Gosztyla ML, Brothers HM, Robinson SR. Alzheimer's amyloid-beta is an antimicrobial peptide: a review of the evidence. *J Alzheimers Dis.* 2018 March;62(4):1495-1506. doi: 10.3233/JAD-171133.
44. Bourgade K, Dupuis G, Frost EH, Fülöp T. Anti-viral properties of amyloid-beta peptides. *J Alzheimers Dis.* 2016 Oct;54(3):859-878. doi: 10.3233/JAD-160517.
45. Postagno AA. Pathogenesis of Alzheimer's disease. *Inter J Mol Sci.* 2022 Dec;24(1):107. doi: 10.3390/ijms24010107.
46. Gonzalez-Ortiz F, Turton M, Kac PR, Smirnov D, Premi E, Ghidoni R, et al. Brain-derived Tau: a novel blood-based biomarker for Alzheimer's disease-type neurodegeneration. *Brain.* 2023 March;146(3):1152-1165. doi: 10.1093/brain/awac407.
47. Sarius H, Heneka MT. Microglia in Alzheimer's disease. *J Clin Invest.* 2017 Sept;127(9):3240-3249. doi: 10.1172/JCI90606.
48. Sun N, Victor MB, Park YP, Xiong X, Scannail AN, Leary N, et al. Human microglial state dynamics in Alzheimer' disease progression. *Cell.* 2023 Sept;186(20):4386-4403.e29. doi: 10.1016/j.cell.2023.08.037.
49. Singh D. Astrocytic and microglial cells as the modulators of neuroinflammation in Alzheimer's disease. *J Neuroinflammation.* 2022 Aug;19(1):206. doi: 10.1186/s12974-022-02565-0.
50. Wu Y, Eisel ULM. Microglial-astrocyte communication in Alzheimer's disease. *J Alzheimer's Dis.* 2023 Sept;95(3):785-803. doi: 10.3233/JAD-230199.
51. Hickman S, Izzy S, Sen P, Morsett L, El Khoury J. Microglia in neurodegeneration. *Nature Neurosci.* 2018 Sept;21(10):1359-1369. doi: 10.1038/s41593-018-0242-x.
52. Kwon HS, Koh S-H. Neuroinflammation in neurodegenerative disorders: the role of microglia and astrocytes. *Transl Neurodegener.* 2020 Nov;9(1):42. doi: 10.1186/s40035-020-00221-2.
53. Prater KE, Green KJ, Mamde S, Sun W, Cochoit A, Smith CL, et al. Human microglia show unique transcriptional changes in Alzheimer's disease. *Nature Aging.* 2023 July;3(7):894-907. doi: 10.1038/s43587-023-00424-y.
54. Tonetti MS, Greenwell H, Kornman KS. Staging and grading of periodontitis: framework and proposal of a new classification and case definition. *J Periodontol.* 2018 June;89(Suppl 1):S159-S172. doi: 10.1002/JPER.18-0006.
55. Ide M, Harris M, Stevens A, Sussams R, Hopkins V, Culliford D, et al. Periodontitis and cognitive decline in Alzheimer's disease. *PLoS One.* 2016 March;11(3):e0151081. doi: 10.1371/journal.pone.0151081.
56. Liccardo D, Marzano F, Carraturo F, Guida M, Femminella GD, Bencivenga L, et al. Potential bidirectional relationship between periodontitis and Alzheimer's disease. *Front Physiol.* 2020 July;11:683. doi: 10.3389/fphys.2020.00683.
57. Borsa L, Dubois M, Sacco G, Lupi L. Analysis of the link between periodontal diseases and Alzheimer's disease: a systematic review. *Inter J Environ Res Public Health.* 2021 Sept;18(17):9312. doi: 10.3390/ijerph18179312.
58. Hu X, Zhang J, Qiu Y, Liu Z. Periodontal disease and the risks of Alzheimer's disease and mild cognitive impairment: a systematic review and meta-analysis. *Psychogeriatrics.* 2021 Jul;21(5):813-825. doi: 10.1111/psyg.12743.
59. Li X, Kiprowska M, Kansara T, Kansara P, Li P. Neuroinflammation: a distal consequence of periodontitis. *J Dent Res.* 2022 Nov;101(12):1441-1449. doi: 10.1177/00220345221102084.
60. Ye W, Tao Y, Wang W, Yu Y, Li X. Periodontitis associated with brain function impairment in middle-aged and elderly individuals with normal cognition. *J Periodontol.* 2025 Mar;96(3):290-300. doi: 10.1002/JPER.24-0264.
61. 2024 Alzheimer's disease facts and figures. *Alzheimer's Dement.* 2024 April;20(5):3708-3821. doi: 10.1002/alz.13809.
62. Pihlstrom BL, Bryan S, Michalowicz BS, Johnson NW. Periodontal diseases. *Lancet.* 2005 Nov;366(9499):1809-1820. doi: 10.1016/S0140-6736(05)67728-8.
63. Hajishengallis G, Lamont RJ. Beyond the red complex and into more complexity: the polymicrobial synergy and dysbiosis (PSD) model of periodontal disease etiology. *Mol Oral Microbiol.* 2012 Dec;27(6):409-419. doi: 10.1111/j.2041-1014.2012.00663.x.
64. Silva N, Abusleme L, Bravo D, Dutzan N, Garcia-Sesnich J, Vernal R, et al. Host response mechanisms in periodontal diseases. *J Appl Oral Sci.* 2015 May-Jun;23(3):329-355. doi: 10.1590/1678-775720140259.
65. Pan C, Liu J, Wang H, Song J, Tan L, Zhao H. *Porphyromonas gingivalis* can invade periodontal ligament stem cells. *BMC Microbiol.* 2017 Feb;17(1):38. doi: 10.1186/s12866-017-0950-5.
66. Hajishengallis G, Liang S, Payne MA, Hashim A, Jotwani R, Eskan MA, et al. Low-abundance biofilm species orchestrates periodontal disease through the commensal microbiota and complement. *Cell Host Microbe.* 2011 Nov;10(5):497-506. doi: 10.1016/j.chom.2011.10.006.
67. Hajishengallis G, Darveau RP, Curtis MA. The keystone pathogen hypothesis. *Nature Rev Microbiol.* 2012 Sept;10(10):717-725. doi: 10.1038/nrmiro2873.
68. How KY, Song KP, Chan KG. *Porphyromonas gingivalis*: an overview of periodontopathic pathogen below the gum line. *Front Microbiol.* 2016 Feb;7:53. doi: 10.3389/fmicb.2016.00053.
69. Olsen I, Lambris JD, Hajishengallis G. *Porphyromonas gingivalis* disturbs host-commensal homeostasis by changing complement function. *J Oral Microbiol.* 2017 June;9(1):1340085. doi: 10.1080/20002297.2017.1340085.
70. Lamont RJ, Jenkinson HF. Life below the gum line: pathogenic mechanisms of *Porphyromonas gingivalis*. *Microbiol Mol Biol Rev.* 1998 Dec;62(4):1244-1263. doi: 10.1128/MMBR.62.4.1244-1263.1998.
71. Hajishengallis G. Immunomicrobial pathogenesis of periodontitis: keystones, pathobionts, and host response. *Trends Immunol.* 2014 Jan;35(1):3-11. doi: 10.1016/j.it.2013.09.001.

72. Holt SC, Kesavalu L, Walker S, Genco CA. Virulence factors of *Porphyromonas gingivalis*. *Periodontol* 2000. 1999 June;20(1):168-238. doi: 10.1111/j.1600-0757.1999.tb00162.x.
73. Xu W, Zhou W, Wang H, Liang S. Roles of *Porphyromonas gingivalis* and its virulence vectors in periodontitis. *Adv Protein Chem Struct Biol*. 2020 Jan;120:45-84. doi: 10.1016/bs.apcsb.2019.12.001.
74. Li Y-Y, Cai Q, Li B-S, Qiao S-W, Jiang J-Y, Wang D, et al. The effect of *Porphyromonas gingivalis* lipopolysaccharide on the pyroptosis of gingival fibroblasts. *Inflammation*. 2021 Jun;44(3):846-858. doi: 10.1007/s10753-020-01379-7.
75. Cheng R, Wen Liu W, Zhang R, Feng Y, Bhowmick NA, Hu T. *Porphyromonas gingivalis*-derived lipopolysaccharide combines hypoxia to induce caspase-1 activation in periodontitis. *Front Cell Infect Microbiol*. 2017 Nov 14;7:474. doi: 10.3389/fcimb.2017.00474.
76. Enersen M, Nakano K, Amano A. *Porphyromonas gingivalis* fimbriae. *J Oral Microbiol*. 2013 May;5:20265. doi: 10.3402/jom.v5i0.20265.
77. Chen WA, Dou Y, Fletcher HM, Boskovic DS. Local and systemic effects of *Porphyromonas gingivalis* infection. *Microorganisms*. 2023 Feb;11(2):470. doi: 10.3390/microorganisms11020470.
78. Tjokro NO, Rocco CJ, Priyadarshini R, Davey ME, Goodman SD. A biochemical analysis of the interaction of *Porphyromonas gingivalis* HU PG0121 protein with DNA. *PLoS One*. 2014 Mar;9(3):e93266. doi: 10.1371/journal.pone.0093266.
79. d'Empaire G, Baer MT, Frank C, Gibson 3rd FC. The K1 serotype capsular polysaccharide of *Porphyromonas gingivalis* elicits chemokine production from murine macrophages that facilitates cell migration. *Infect Immun*. 2006 Nov;74(11):6236-6243. doi: 10.1128/IAI.00519-06.
80. Okamura H, Hirota K, Yoshida K, Weng Y, He Y, Shiotsu N, et al. Outer membrane vesicles of *Porphyromonas gingivalis*: novel communication tool and strategy. *Jpn Dent Sci Rev*. 2021 Nov;57:138-146. doi: 10.1016/j.jdsr.2021.07.003.
81. Chuang W-C, Yang C-N, Wang H-W, Lin S-K, Yu C-C, Syu J-H, et al. The mechanisms of *Porphyromonas gingivalis*-derived outer membrane vesicles-induced neurotoxicity and microglia activation. *J Dent Sci*. 2024 July;19(3):1434-1442. doi: 10.1016/j.jds.2024.04.002.
82. Uemura Y, Hiroshima Y, Tada A, Murakami K, Yoshida K, Inagaki Y, et al. *Porphyromonas gingivalis* outer membrane vesicles stimulate gingival epithelial cells to induce pro-inflammatory cytokines via the MAPK and STING pathways. *Biomedicine*. 2022 Oct;10(10):2643. doi: 10.3390/biomedicine10102643.
83. Veith PD, Chen Y-Y, Gorasia DG, Chen D, Glew MD, O'Brien-Simpson NM, et al. *Porphyromonas gingivalis* outer membrane vesicles exclusively contain outer membrane and periplasmic proteins and carry a cargo enriched with virulence factors. *J Proteome Res*. 2014 May;13(5):2420-2432. doi: 10.1021/pr401227e.
84. de Diego I, Veillard F, Sztukowska MN, Guevara T, Potempa B, Pomowski A, et al. Structure and mechanism of cysteine peptidase gingipain K (Kgp), a major virulence factor of *Porphyromonas gingivalis* in periodontitis. *J Biol Chem*. 2014 Nov;289(46):32291-32302. doi: 10.1074/jbc.M114.602052.
85. Bostanci N, Belibasakis GN. *Porphyromonas gingivalis*: an invasive and evasive opportunistic oral pathogen. *FEMS Microbiol Lett*. 2012 Aug;333(1):1-9. doi: 10.1111/j.1574-6968.2012.02579.x.
86. Kadowaki T. Enzymatic characteristics and activities of gingipains from *Porphyromonas gingivalis*. *Methods Mol Biol*. 2021;2210:97-112. doi: 10.1007/978-1-0716-0939-2_10.
87. Imamura T. The role of gingipains in the pathogenesis of periodontal disease. *J Periodontol*. 2003 Jan;74(1):111-118. doi: 10.1902/jop.2003.74.1.111.
88. Houle M-A, Grenier D, Plamondon P, Nakayama K. The collagenase activity of *Porphyromonas gingivalis* is due to Arg-gingipain. *FEMS Microbiol Lett*. 2003 April;221(2):181-185. doi: 10.1016/S0378-1097(03)00178-2.
89. Zhou J, Windsor J. *Porphyromonas gingivalis* affects host collagen degradation by affecting expression, activation, and inhibition of matrix metalloproteinases. *J Periodontal Res*. 2006 Feb;41(1):47-54. doi: 10.1111/j.1600-0765.2005.00835.x.
90. Travis J, Pike R, Imamura T, Potempa J. *Porphyromonas gingivalis* proteinases as virulence factors in the development of periodontitis. *J Periodontal Res*. 1997 Jan;32(1 Pt 2):120-125. doi: 10.1111/j.1600-0765.1997.tb01392.x.
91. Sochalska M, Potempa J. Manipulation of neutrophils by *Porphyromonas gingivalis* in the development of periodontitis. *Front Cell Infect Microbiol*. 2017 May;7:197. doi: 10.3389/fcimb.2017.00197.
92. Singhrao SK, Harding A. Is Alzheimer's disease a polymicrobial host microbiome dysbiosis? *Expert Rev Anti Infect Ther*. 2020 April;18(4):275-277. doi: 10.1080/14787210.2020.1729741.
93. Sedghi L, DiMassa V, Harrington A, Lynch SV, Kapila YL. The oral microbiome: role of key organisms and complex networks in oral health and disease. *Periodontol* 2000. 2021 Oct;87(1):107-131. doi: 10.1111/prd.12393.
94. Oliveira RRDS, Fermiano D, Feres M, Figueiredo LC, Teles FRF, Soares GMS, et al. Levels of candidate periodontal pathogens in subgingival biofilm. *J Dent Res*. 2016 June;95(6):711-718. doi: 10.1177/0022034516634619.
95. Antezack A, Etchecopar-Etchart D, La Scola B, Monnet-Corti V. New putative periodontopathogens and periodontal health-associated species: a systematic review. *J Periodontal Res*. 2023 Oct;58(5):893-906. doi: 10.1111/jre.13173.
96. Wang RP-H, Huang J, Chan KKY, Leung WK, Goto T, Ho Y-S, et al. IL-1 β and TNF- α play an important role in modulating the risk of periodontitis and Alzheimer's disease. *J Neuroinflammation*. 2023 March;20(1):71. doi: 10.1186/s12974-023-02747-4.

97. Villar A, Paladini S, Cossatis J. Periodontal disease and Alzheimer's: insights from a systemic literature network analysis. *J Prev Alzheimer's Dis.* 2024 April;11(4):1148-1165. doi: 10.14283/jpad.2024.79.
98. Olsen I. Porphyromonas gingivalis-induced neuroinflammation in Alzheimer's disease. *Front Neurosci.* 2021 Oct;15:691016. doi: 10.3389/fnins.2021.691016.
99. Kanagasalingam S, Chukkapalli SS, Singhrao SK. Porphyromonas gingivalis is a strong risk factor for Alzheimer's disease. *J Alzheimer's Dis Rep.* 2020 Dec;4(4):501-511. doi: 10.3233/ADR-200250.
100. Haditsch U, Roth T, Rodriguez L, Hancock S, Cecere T, Nguyen M, et al. Alzheimer's disease-like neurodegeneration in Porphyromonas gingivalis infected neurons with persistent expression of active gingipains. *J Alzheimer's Dis.* 2020 April;75(4):1361-1376. doi: 10.3233/JAD-200393.
101. Haraszthy VI, Zambon JJ, Trevisan M, Zeid M, Genco RJ. Identification of periodontal pathogens in atheromatous plaques. *J Periodontol.* 2000 Oct;71(10):1554-1560. doi: 10.1902/jop.2000.71.10.1554.
102. Dominy SS, Lynch C, Ermini F, Benedyk M, Marczyk A, Konradi A, et al. Porphyromonas gingivalis in Alzheimer's disease brains: evidence for disease causation and treatment with small-molecule inhibitors. *Sci Adv.* 2019 Jan;5(1):eaau3333. doi: 10.1126/sciadv.aau3333.
103. Jungbauer G, Stähli A, Zhu X, Alberi LA, Sculean A, Eick S. Periodontal microorganisms and Alzheimer disease – a causative relationship? *Periodontol* 2000. 2022 Jun;89(1):59-82. doi: 10.1111/prd.12429.
104. Jones TB, Chu P, Wilkey B, Lynch L, Jentarra G. Regional differences in microbial infiltration of brain tissue from Alzheimer's disease patients and control individuals. *Brain Sci.* 2024 Jul;14(7):677. doi: 10.3390/brainsci14070677.
105. Lim SL, Rodriguez-Ortiz CJ, Kitazawa M. Infection, systemic inflammation, and Alzheimer's disease. *Microbes Infect.* 2015 Aug;17(8):549-556. doi: 10.1016/j.micinf.2015.04.004.
106. Bu X-L, Yao X-Q, Jiao S-S, Feng F, Liu Y-H, Xiang Y, et al. A study on the association between infectious burden and Alzheimer's disease. *Eur J Neurol.* 2015 Dec;22(12):1519-1525. doi: 10.1111/ene.12477.
107. Zhao M, Wang Y, Shen Y, Wei C, Zhang G, Sun L. A review of the roles of pathogens in Alzheimer's disease. *Front Neurosci.* 2024 Aug;18:1439055. doi: 10.3389/fnins.2024.1439055.
108. Li R, Wang J, Xiong W, et al. The oral-brain axis: can periodontal pathogens trigger the onset and progression of Alzheimer's disease? *Front Microbiol.* 2024 Jan;15:1358179. doi: 10.3389/fmicb.2024.1358179.
109. Cichońska D, Mazuś M, Kusiak A. Recent aspects of periodontitis and Alzheimer's disease: a narrative review. *Inter J Mol Sci.* 2024 Feb;25(5):2612. doi: 10.3390/ijms25052612.
110. Tang D, Sun C, Yang J, Fan L, Wang Y. Advances in the study of the pathology and treatment of Alzheimer's disease and its association with periodontitis. *Life (Basel).* 2023 Nov;13(11):2203. doi: 10.3390/life13112203.
111. de Jongh CA, de Vries TJ, Bikker FJ, Gibbs S, Krom BP. Mechanisms of Porphyromonas gingivalis to translocate over the oral mucosa and other tissue barriers. *J Oral Microbiol.* 2023 April;15(1):2205291. doi: 10.1080/20002297.2023.2205291.
112. Olsen I, Singhrao SK. Interaction between genetic factors, Porphyromonas gingivalis and microglia to promote Alzheimer's disease. *J Oral Microbiol.* 2020 Sept;12(1):1820834. doi: 10.1080/20002297.2020.1820834.
113. Abt E, Kumar S, Weyant RJ. Periodontal disease and medical maladies: what do we really know? *J Am Dent Assoc.* 2022 Jan;153(1):9-13. doi: 10.1016/j.adaj.2021.08.014.
114. Huang D, Wang Y-Y, Li B-H, Wu L, Xie W-Z, Zhou X, et al. Association between periodontal disease and systemic diseases: a cross-sectional analysis of current evidence. *Mil Med Res.* 2024 Dec 4;11(1):74. doi: 10.1186/s40779-024-00583-y.
115. Gao C, Larvin H, Bishop DT, Bunce D, Pavitt S, Wu J, et al. Oral diseases are associated with cognitive function in adults over 60 years old. *Oral Dis.* 2024 Jul;30(5):3480-3488. doi: 10.1111/odi.14757.
116. Chen L, Li X, Liu J, Hou Z, Wei Y, Chen M, et al. Distinctive subgingival microbial signatures in older adults with different levels of cognitive function. *J Clin Periodontol.* 2024 Aug;51(8):1066-1080. doi: 10.1111/jcpe.13997.